

AC Plant Keratin PF Efficacy Data

Code: 20624PF
INCI Name: Hydrolyzed Corn Protein & Hydrolyzed Wheat Protein & Hydrolyzed Soy Protein
CAS #: 100209-41-4 & 100209-45-8 & 68607-88-5
EINECS #: 309-349-6 & 309-353-8 & 271-770-5

Name of Study	Results
<p>ORAC Assay</p>	<p>As shown in figure 1, AC Plant Keratin PF exhibited positive results by increasing cell metabolism. The increase in fluorescent signal indicates an increase in cellular metabolism and viability post AC Plant Keratin PF treatment. For these reasons, we can assume AC Plant Keratin PF is suitable for cosmetic applications designed to increase cell viability and metabolism.</p>
<p>Cellular Viability Assay</p>	<p>As shown in figure 1, AC Plant Keratin PF exhibited antioxidant activity comparable to 200µM Trolox®. The antioxidant capacity of AC Plant Keratin PF increased as the concentration increased. As a result we can assure that its ability to minimize oxidative stress is dose dependent. With the present study we can confirm that this unique ingredient is capable of providing antioxidant benefits when added to cosmetic applications.</p>



Oxygen Radical Absorbance Capacity (ORAC) Assay

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Tradename: AC Plant Keratin PF

Code: 20624PF

CAS #: 100209-41-4 & 70084-87-6 (or) 94350-06-8 & 68607-88-5

Test Request Form #: 613

Lot #: 30147

Sponsor: *Active Concepts, LLC; 107 Technology Drive Lincolnton, NC 28092*

Study Director: *Erica Segura*

Principle Investigator: *Meghan Darley*

Test Performed:

Oxygen Radical Absorbance Capacity (ORAC)

Introduction

Reactive oxygen species (ROS) are generated by normal cellular processes, environmental stresses, and UV irradiation. ROS are detrimental to cellular structures and functional molecules (i.e DNA, proteins, lipids) as they act as strong oxidizing agents or free radicals. The oxygen radical absorbance capacity (ORAC) assay is a standard method used to assess antioxidant capacity of physiological fluids, foods, beverages, and natural products. The assay quantitatively measures a sample's ability to quench free radicals that have the potential to react with and damage cellular components.

Oxygen Radical Absorbance Capacity (ORAC) assay was conducted to assess the antioxidant capacity of **AC Plant Keratin PF**.

Assay Principle

This assay is based upon the effect of peroxy radicals generated from the thermal decomposition of 2, 2'-azobis-2-methyl-propanimidamide dihydrochloride (AAPH) on the signal intensity from the fluorescent probe, fluorescein, in the presence of an oxygen radical absorbing substance. The degree of change is indicative of the amount of radical damage and the presence of antioxidants results in an inhibition in the free radical damage to the fluorescein. The antioxidant protection of the sample can be calculated by comparing it to a set of known standards. Trolox®, a water soluble vitamin E analog, with known antioxidant capabilities is used in this ORAC assay as the standard for measuring the antioxidant capacity of unknown substances. ORAC values, expressed in μM of Trolox® equivalents (TE), are calculated using the area under the curves (AUC) of the test product, Trolox®, and the control materials. Trolox equivalency is used as the benchmark for antioxidant capacity of mixtures since it is difficult to measure individual components.

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Materials

- A. Equipment:** Synergy H1 Microplate reader (BioTek Instruments, Winooski, VT); Gen5 software (BioTek Instruments, Winooski, VT); Pipettes
- B. Buffers:** 75mM Potassium Phosphate (pH 7.4); Deionized H₂O
- C. Reagents:** 2,2'-Azobis(2-methylpropanimidine) dihydrochloride (AAPH) (153mM); 6-Hydroxy-2,5,7,8-tetramethylchromane-2-carboxylic acid (Trolox®); Fluorescein Sodium Salt (4nM)
- D. Preparation:** Pre-heat (37°C) Synergy H1 Microplate reader; Prepare Trolox® standards, sample dilutions, fluorescein solution, and AAPH.
- E. Microtitre Plates:** Corning 96 Well Black Side/Clear Bottom Microplates

Methods

Solutions of **AC Plant Keratin PF** and Trolox® (positive control) were prepared in 75mM potassium phosphate buffer. Materials were prepared at three different concentrations/dilutions. Trolox® was used as a reference for antioxidant capacity and prepared at a concentrations ranging from 12.5µM to 200µM in 75mM potassium phosphate buffer.

For the ORAC assay, 25µL of test material and Trolox® were combined with 150µL of fluorescein in 75mM potassium phosphate buffer and incubated in the Synergy HT Microplate reader at 37°C for 30 minutes. At the end of the incubation period, 25µL of AAPH were pipetted into each well. Fluorescent measurements were then taken every 2 minutes for approximately 2 hours at an excitation wavelength of 485nm and an emissions wavelength of 520nm.

The AUC and Net AUC values of the standards and samples were determined using Gen5 2.0 Data Reduction Software using the below equations:

$$AUC = 0.5 + \frac{R2}{R1} + \frac{R3}{R1} + \frac{R4}{R1} + \dots + \frac{Rn}{R1} \rightarrow \text{Where } R \text{ is fluorescence reading}$$

$$\text{Net AUC} = AUC_{\text{sample}} - AUC_{\text{blank}}$$

The standard curve was obtained by plotting the Net AUC of different Trolox® concentrations against their concentration. ORAC values of samples were then calculated automatically using the Gen5 software to interpolate the sample's Net AUC values against the Trolox® standard curve. ORAC measurements for the test material were expressed in micro moles Trolox® equivalents (µMTE), where 1 ORAC unit is equal to 1 µMTE.

ORAC values are also calculated in Units/milliliter (U/mL). The equation used for the calculation is shown below:

$$ORAC (U/mL) = (50 \times \text{Dilution Factor}) \times \left(\frac{AUC_{\text{Sample}} - AUC_{\text{Blank}}}{AUC_{\text{Trolox}} - AUC_{\text{Blank}}} \right)$$

Results

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AC Plant Keratin PF began exhibiting antioxidant activity at a 0.1% concentration.

The ORAC value expressed in U/mL for 0.5% AC Plant Keratin PF is 9174.13.

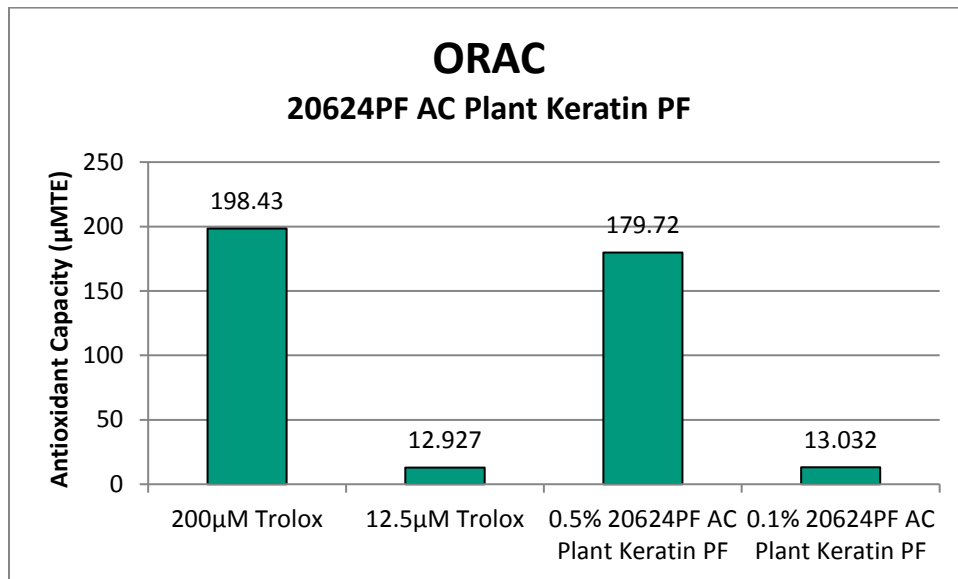


Figure 1: Antioxidant capacities

Discussion

As shown in figure 1, AC Plant Keratin PF exhibited antioxidant activity comparable to 200µM Trolox®. The antioxidant capacity of AC Plant Keratin PF increased as the concentration increased. As a result we can assure that its ability to minimize oxidative stress is dose dependent. With the present study we can confirm that this unique ingredient is capable of providing antioxidant benefits when added to cosmetic applications.

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Cellular Viability Assay Analysis

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Sponsor: *Active Concepts, LLC; 107 Technology Drive Lincolnton, NC 28092*

Study Director: *Erica Segura*

Principle Investigator: *Meghan Darley*

Test Performed:

Cellular Viability Assay

Introduction

The cellular viability assay is useful for quantitatively measuring cell-mediated cytotoxicity, cell proliferation and mitochondrial metabolic activity. Increased metabolism in a cell indicates ample cellular respiration and adenosine triphosphate (ATP) production. ATP is the molecular energy of cells and is required in basic cell function and signal transduction. A decrease in ATP levels indicates cytotoxicity and decreased cell function while an increase in ATP levels indicates healthy cells.

The cellular viability assay was conducted to assess the ability of **AC Plant Keratin PF** to increase cellular metabolic activity in cultured dermal fibroblasts.

Assay Principle

The assay utilizes a nonfluorescent dye, resazurin, which is converted to a fluorescent dye, resorufin, in response to chemical reduction of growth medium from cell growth and by respiring mitochondria. Healthy cells that are in a proliferative state will be able to easily convert resazurin into resorufin without harming the cells. This method is a more sensitive assay than other commonly used mitochondrial reductase dyes such as MTT. An increase in the signal generated by resazurin-conversion is indicative of a proliferative cellular state.

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Cellular Viability Assay Analysis

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Materials

- A. Kit:** Cellular Senescence Assay Kit (Chemicon® International; KAA002)
- B. Incubation Conditions:** 37°C at 5% CO₂ and 95% relative humidity (RH)
- C. Equipment:** Forma humidified incubator; ESCO biosafety laminar flow hood; Light microscope; Pipettes
- D. Cell Line:** Normal Human Dermal Fibroblasts (NHDF) (Lonza; CC-2511)
- E. Media/Buffers:** Dulbecco's Modified Eagle Medium (DMEM); Penicillin-Streptomycin (50U-50mg/mL); Fetal Bovine Serum (FBS); Phosphate Buffered Saline (PBS)
- F. Culture Plate:** Falcon flat bottom 6-well tissue culture treated plates
- G. Reagents:** 5-bromo-4-chloro-indolyl-β-D-galactopyranoside (X-gal)
- H. Other:** Sterile disposable pipette tips; wash bottles; 15mL conical tubes, 1.5mL microcentrifuge tubes

Methods

Human dermal fibroblasts were seeded into 96-well tissue culture plates and allowed to grow to confluency in complete DMEM. A 10-fold serial dilution was performed resulting in **AC Plant Keratin PF** concentrations on 1%, 0.1%, and 0.01% in complete DMEM and incubated with fibroblasts for 24 hours.

Ten microliters of viability reagent was added to 90μL of cell culture media in culture wells.

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Results

The data obtained from this study met criteria for a valid assay and the controls performed as anticipated.

AC Plant Keratin PF exhibited positive effects on cellular metabolism compared to the control.

Cellular metabolism results are expressed as a percentage of the control.

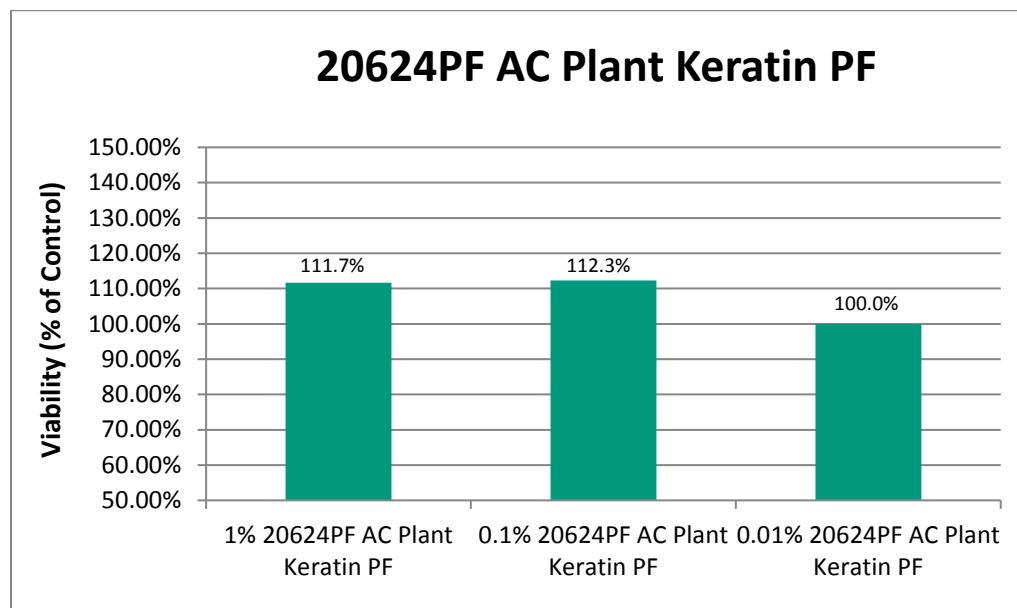


Figure 1: Cellular Metabolism of **AC Plant Keratin PF** -treated fibroblasts expressed in terms of percent of control.

Discussion

As shown in figure 1, **AC Plant Keratin PF** exhibited positive results by increasing cell metabolism. The increase in fluorescent signal indicates an increase in cellular metabolism and viability post **AC Plant Keratin PF** treatment. For these reasons, we can assume **AC Plant Keratin PF** is suitable for cosmetic applications designed to increase cell viability and metabolism.