

IL-6 ELISA Analysis

info@activeconceptsllc.com • Phone: +1-704-276-7100 • Fax: +1-704-276-7101

Tradename: ABS Willow Bark Extract

Code: 10200

CAS #: 84650-64-6

Test Request Form #: 1113

Sponsor: Active Concepts, LLC; 107 Technology Drive Lincolnton, NC 28092

Study Director: Erica Segura

Principle Investigator: Meghan Darley

Test Performed:

Interleukin (IL)-6 Enzyme-Linked Immunosorbent Assay (ELISA)

Introduction

Interleukin-6 is a proinflammatory cytokine known to play an active role in inflammation, immunology, bone metabolism, reproduction, arthritis, neoplasia, and aging. IL-6 signals through the nuclear factor-kappa B (NF- κ B) pathway that results in the transcription of inflammatory mediators, including matrix metalloproteinase-1 (MMP-1). MMP's are responsible for breaking down the extracellular matrix and collagen in the skin leading to wrinkles, fine lines, and loss of skin elasticity. Reducing the level of IL-6 and other inflammatory mediators is believed to slow down degradation of the skin matrix and, possibly, stimulate its replenishment.

Interleukin-6 ELISA was conducted to assess the changes in IL-6 levels in **ABS Willow Bark Extract**-treated *in vitro* cultured human dermal fibroblasts.

Assay Principle

This ELISA utilizes a colorimetric reaction employing antibodies with antigen specificity to human IL-6. Monoclonal antibodies specific for IL-6 epitopes are coated on a microtiter plate. In positive samples, IL-6 will bind to these antibodies and are tagged a second time with another IL-6-specific antibody labeled with horseradish peroxidase (HRP). The addition of the chromagen solution, containing 3,3',5,5'-tetramethylbenzidine, provides the colorimetric reaction with HRP that is quantitated through optical density (OD) readings on a microplate spectrometer. The standard curve provides a reference from the OD readings for the amount of collagen in each sample.



F. Culture Plate:

IL-6 ELISA Analysis

info@activeconceptsllc.com • Phone: +1-704-276-7100 • Fax: +1-704-276-7101

Materials

A. Kit: IL-6 ELISA Kit (Biosource; KAC1261)

B. Incubation Conditions: 37°C at 5% CO₂ and 95% relative humidity (RH)

C. Equipment: Forma humidified incubator; ESCO biosafety laminar flow

hood; Microplate Reader; Pipettes

D. Cell Line: Normal Human Dermal Fibroblasts (NHDF) (Lonza; CC-2511)
E. Media/Buffers: Dulbecco's Modified Eagle Medium (DMEM); Penicillin-

Streptomycin (50U-50mg/mL); Fetal Bovine Serum (FBS);

Phosphate Buffered Saline (PBS); Amphotericin (45pg/mL)

Falcon flat bottom 12-well tissue culture treated plates

G. Reagents: Lipopolysaccharide (LPS) (1µg/mL); Solution A (human

plasma with preservatives);

Solution B (buffer with preservative)

H. Other: Sterile disposable pipette tips; wash bottles

Methods

Human dermal fibroblasts were seeded into 12-well tissue culture plates and allowed to grow to confluency in complete DMEM. 1%, 0.1%, 0.01% concentrations of **ABS Willow Bark Extract** were added to complete DMEM containing 1µg/mL LPS and incubated with fibroblasts for 24 hours. Complete media containing 1µg/mL LPS was used as the positive controls and complete DMEM was used a negative control.

Standards were prepared in concentrations ranging from 0pg/mL to 2476pg/mL. $50\mu L$ of Solution B was added to wells for standards and assay controls and $50\mu L$ of Solution A was added to experiment wells. $100\mu L$ of standards, controls, and samples were added to appropriate wells. After a one hour incubation at room temperature and washing, $50\mu L$ Solution A and $100\mu L$ anti-IL-6 conjugate was added to all wells. Following a one hour incubation and washing, $100~\mu L$ chromagen solution was added for the colorimetric reaction. One-hundred μL stop solution was added to stop the reaction after 15 minutes. The optical density was read at 450nm on the Synergy HT Microplate Reader.

A standard curve was created by reducing the data and generating a linear curve fit. The IL-6 concentration of **ABS Willow Bark Extract** treated-fibroblasts was determined by extrapolation from the standard curve and expressed in pg/mL.



IL-6 ELISA Analysis

info@activeconceptsllc.com • Phone: +1-704-276-7100 • Fax: +1-704-276-7101

Results

The data obtained from this study met criteria for a valid assay and the positive and negative controls performed as anticipated.

ABS Willow Bark Extract, at a concentration of 1% was able to decrease IL-6 production compared to our positive control.

IL-6 levels are expressed by the following formula:

 $= Average_{IL-6\;Concentrations} \times Dilution\;Factor$

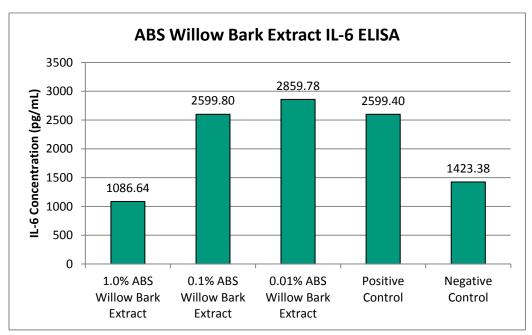


Figure 1: ABS Wilow Bark Extract-treated fibroblasts IL-6 concentrations



IL-6 ELISA Analysis

info@activeconceptsllc.com • Phone: +1-704-276-7100 • Fax: +1-704-276-7101

IL-6 production percent decrease is calculated by the following formula:

$$= \frac{Positive \; Control_{\; Avg.Concentration} - \; Sample_{\; Avg.Concentration}}{Positive \; Control_{\; Avg.Concentration}} \times 100$$

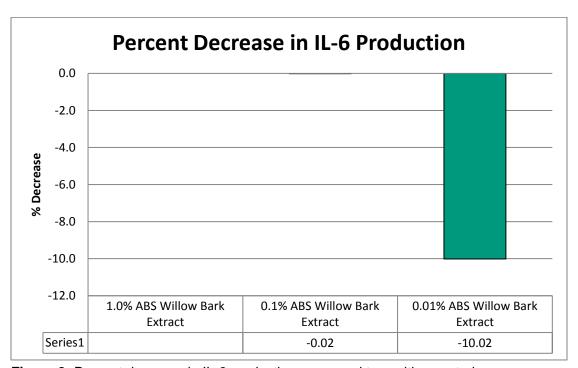


Figure 2: Percent decrease in IL-6 production compared to positive control

Discussion

As shown in figure 1, **ABS Willow Bark Extract** exhibited anti-inflammatory effects on LPS-treated fibroblasts. As expected, the changes in IL-6 production using **ABS Willow Bark Extract** appears to be dose dependent. This decrease in IL-6 production indicates a reduced inflammatory environment which could decrease the signs of aging and reduce the formation of fine lines and wrinkles. For these reasons, we can assume **ABS Willow Bark Extract** is suitable for cosmetic applications designed to provide soothing and anti-aging properties.