



**Tradename:** AC Biopolymer Chia PF

**Code:** 21005PF

**CAS #:** 61788-47-4 & 93384-40-8

**Test Request Form #:** 3627

**Lot #:** NC170710-F

**Sponsor:** *Active Concepts, LLC; 107 Technology Drive Lincolnton, NC 28092*

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**Test Performed:**

OECD TG 442D: *In Vitro* Skin Sensitization *ARE-Nrf2 Luciferase Test Method*

## Introduction

Skin sensitization refers to an allergic response following skin contact with the tested chemical, as defined by the United Nations Globally Harmonized System of Classification and Labelling of Chemicals<sup>1</sup>. Substances are classified as skin sensitizers if there is evidence in humans that the substance can lead to sensitization by skin contact or positive results from appropriate tests, both *in vivo* and *in vitro*. Utilization of the KeratinoSens™ cell line allows for valid *in vitro* testing for skin sensitization.

This assay was conducted to determine skin sensitization potential of **AC Biopolymer Chia PF** in accordance with the UN GHS.

## Assay Principle

The ARE-Nrf2 luciferase test method addresses the induction of genes that are regulated by antioxidant response elements (ARE) by skin sensitizers. The Keap1-Nrf2-ARE pathways have been shown to be major regulator of cytoprotective responses to oxidative stress or electrophilic compounds. These pathways are also known to be involved in the cellular processes in skin sensitization. Small electrophilic substances such as skin sensitizers can act on the sensor protein Keap1 (Kelch-like ECH-associated protein 1), by covalent modification of its cysteine residue, resulting in its dissociation from the transcription factor Nrf2 (nuclear factor-erythroid 2-related factor 2). The dissociated Nrf2 can then activate ARE-dependent genes such as those coding for phase II detoxifying enzymes.

The skin sensitization assay utilizes the KeratinoSens™ method which uses an immortalized adherent human keratinocyte cell line (HaCaT cell line) that has been transfected with a selectable plasmid to quantify luciferase gene induction as a measure of activation of Keap1-Nrf2-antioxidant/electrophile response element (ARE). This test method has been validated by independent peer review by the EURL-ECVAM. The addition of a luciferin containing reagent to the cells will react with the luciferase produced in the cell resulting in luminescence which can be quantified with a luminometer.

1. United Nations (UN) (2013). Globally Harmonized System of Classification and Labelling of Chemicals (GHS), Fifth revised edition, UN New York and Geneva, 2013

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## Materials

- A. Incubation Conditions:** 37 °C at 5% CO<sub>2</sub> and 95% relative humidity (RH)
- B. Equipment:** Humidified incubator; Biosafety laminar flow hood; Microplate Reader; Pipettes
- C. Cell Line:** KeratinoSens™ by Givaudan Schweiz AG
- D. Media/Buffers:** Dulbecco's Modified Eagle Medium (DMEM); Fetal Bovine Serum (FBS); Phosphate Buffered Saline (PBS); Geneticin
- E. Culture Plate:** Flat bottom 96-well tissue culture treated plates
- F. Reagents:** Dimethyl Sulfoxide (DMSO); Cinnamic Aldehyde; ONE-Glo Reagent; 3-(4,5-dimethylthiazol-2-yl)-2,5-diphenyltetrazolium bromide (MTT); sodium lauryl sulfate (SLS)
- G. Other:** Sterile disposable pipette tips; wash bottles

## Methods

KeratinoSens™ were into seeded four 96-well tissue culture plates and allowed to grow to 80 – 90% confluency in DMEM containing 10% FBS and 500µg/mL G418 geneticin. Twelve test concentrations of **AC Biopolymer Chia PF** were prepared in DMSO with a concentration range from 0.98 - 2000 µM. These 12 concentrations were assayed in triplicate in 2 independently performed experiments. The positive control was cinnamic aldehyde for which a series of 5 concentrations prepared in DMSO had final test concentrations of 4 – 64 µM. The negative control was a 1% test concentration of DMSO.

24 hour post KeratinoSens™ seeding, the culture media was removed and replaced with fresh media containing 10% FBS without G418 geneticin. 50 µL of the above described test concentrations was added to the appropriate wells. The treated plates were then incubated for 48 hours at 37 °C in the presence of 5% CO<sub>2</sub> and 95% relative humidity. After treatment incubation was complete the media was removed and the wells were washed with PBS 3 times.

One of the four plates was used for a cytotoxicity endpoint, where MTT was added to the wells and incubated for 4 hours at 37 °C in the presence of 5% CO<sub>2</sub>. SLS was then added to the wells and incubated overnight at room temperature. A spectrometer measured the absorbance at 570 nm. The absorbance values (optical density) were then used to determine the viability of each well by comparing the optical density of each test material treated well to that of the solvent control wells to determine the IC<sub>50</sub> and IC<sub>30</sub> values.

The remaining 3 plates were used in the luciferase induction endpoint of the assay. 100 µL of Promega's ONE-Glo Reagent was added to 100 µL of fresh media containing 10% FBS without geneticin. Cells were incubated for 5 minutes to induce cell lysis and release luciferin into the media. Plates were read with a luminometer and EC<sub>1.5</sub> and maximum response (I<sub>max</sub>) values were obtained.

## Data and Reporting

### Acceptance Criteria:

1. Gene induction obtained with the positive control, cinnamic aldehyde, should be statistically significant above the threshold of 1.5 in at least one of the tested concentrations (from 4 to 64  $\mu\text{M}$ ).
2. The EC<sub>1.5</sub> value should be within two standard deviations of the historical mean and the average induction in the three replicates for cinnamic aldehyde at 64  $\mu\text{M}$  should be between 2 and 8.
3. The average coefficient of variability of the luminescence reading for the negative (solvent) control DMSO should be below 20% in each experiment.

A KeratinoSens™ prediction is considered positive if the following conditions are met:

1. The I<sub>max</sub> is higher than 1.5-fold and statistically significantly higher as compared to the solvent (negative) control
2. The cellular viability is higher than 70% at the lowest concentration with a gene induction above 1.5 fold (i.e., at the EC<sub>1.5</sub> determining concentration)
3. The EC<sub>1.5</sub> value is less than 1000  $\mu\text{M}$  (or < 200  $\mu\text{g/ml}$  for test chemicals with no defined MW)
4. There is an apparent overall dose-response for luciferase induction

## Results

Compound	Classification	EC <sub>1.5</sub> ( $\mu\text{M}$ )	IC <sub>50</sub>	I <sub>max</sub>
Cinnamic aldehyde	Sensitizer	19	289.19 $\mu\text{M}$	32.1
DMSO	Non-Sensitizer	No Induction	243.24 $\mu\text{M}$	0.19
<b>AC Biopolymer Chia PF</b>	Non-Sensitizer	No Induction	> 1000 $\mu\text{M}$	0.36

Table 1: Overview of KeratinoSens™ Assay Results (I<sub>max</sub> equals the average induction values Fig.1)

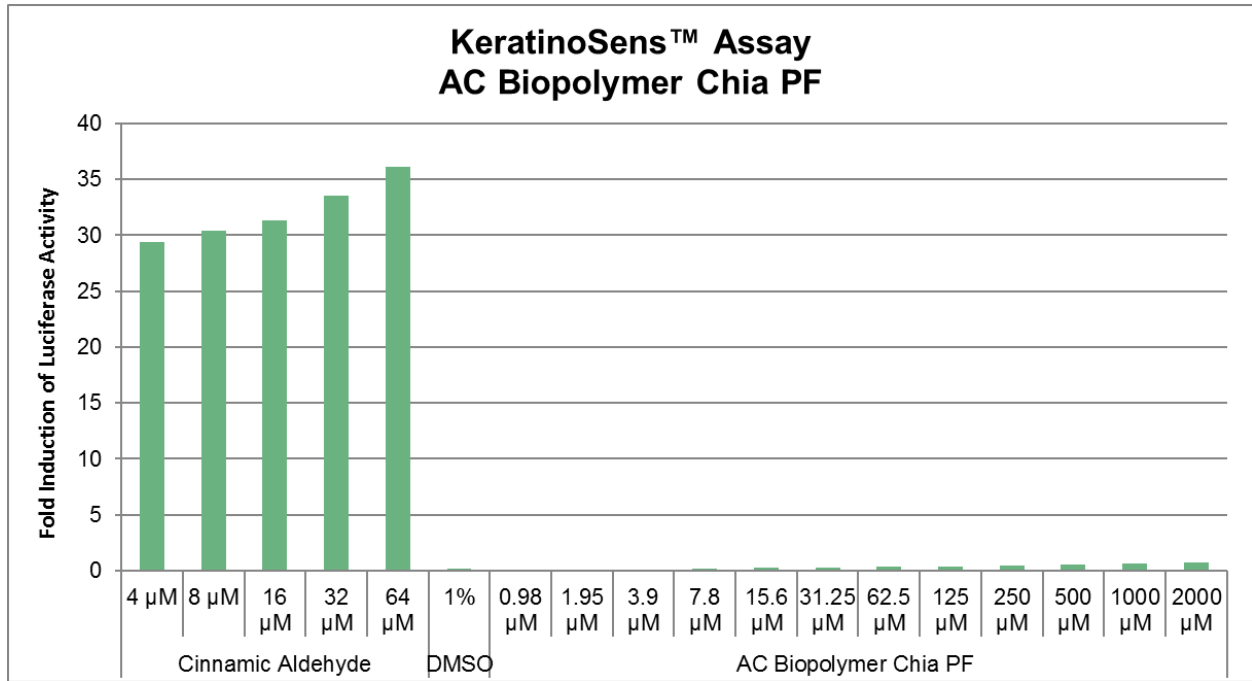


Figure 1: Fold Induction of Luciferase

### Discussion

As shown in the results, **AC Biopolymer Chia PF (21005PF)** was not predicted to be a skin sensitizer based on the KeratinoSens™ ARE-Nrf2 Luciferase Test Method as there was not a significant increase in luciferase expression. It can be concluded that **AC Biopolymer Chia PF** can be safely used in cosmetics and personal care products at typical use levels.